## Stereospecific Synthesis of Chiral Acetic Acid from Glycine

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Summary A convenient chemical synthesis of (2R)- $[^{1}H,^{2}H,^{3}H]$ acetic acid from (2R)- $[^{2}H]$ glycine in high (92%) optical yield is described.

The use of chiral acetic acid, first introduced for the solution of biosynthetic problems by Cornforth<sup>1</sup> and Arigoni<sup>2</sup> in 1969, continues apace<sup>3</sup> and a new synthesis has been reported.<sup>4</sup> Our requirement for substantial (1-5 g) quantities of



SCHEME 1. i, HLAD, NAD<sup>+</sup>, 1-[<sup>2</sup>H or <sup>3</sup>H]cyclohexan-1-ol, (78%); ii, propane-1,3-dithiol, Bu<sup>n</sup>Li, D<sub>2</sub>O(TH<sub>2</sub>O), (90%); iii, CuCl<sub>2</sub>, CuO, (86%); iv, HLAD, NAD<sup>+</sup>, EtOH, (80%) or Na<sub>2</sub>S<sub>2</sub>O<sub>4</sub>, H<sub>2</sub>O, (70%); v, TSCl, NaN<sub>3</sub>, (86%); vi, LiAlH<sub>4</sub>, (92%); vii, RuO<sub>4</sub>,<sup>7</sup> (39%); viii, Ac<sub>2</sub>O, pyridine, (94%); ix, O<sub>3</sub>, HCO<sub>2</sub>H, CHCl<sub>3</sub>, H<sub>2</sub>O<sub>2</sub>, (60%);<sup>8</sup> x, hog kidney acylase I, (93%).<sup>8</sup> HLAD = Horse liver alcohol dehydrogenase. Ts=MeC<sub>8</sub>H<sub>4</sub>SO<sub>2</sub>-p.

chiral glycine has led to an improved sequence for the preparation of this species (R or S;  ${}^{2}H$  or  ${}^{3}H$ ) as summarized in Scheme 1, which includes several modifications of literature procedure,<sup>5</sup> and which provides a viable synthetic method to chiral acetate.

Starting from [<sup>1</sup>H]benzaldehyde (1), (4a;  $R^2 = {}^{2}H$  or  ${}^{3}H$ ) is prepared in 78% overall yield whilst (4b;  $R^1 = {}^{2}H$  or  ${}^{3}H$ ) is synthesized from (3;  $R = {}^{2}H$  or  ${}^{3}H$ ) in 80% yield. Compounds (4a, b) are then converted as shown, with one inversion of configuration  $(5 \rightarrow 6)$  into the (R)-(10b) and (S)-(10a)  $[{}^{2}H_{1} \text{ (or }{}^{3}H_{1})]$ glycines  $[ca. 92-96\% {}^{2}H (m/e 76)$  by mass spectrometry], respectively. The absolute configuration of each <sup>2</sup>H species was established by comparison of the o.r.d. curves with the published data.<sup>6</sup>

A sample of (R)-[<sup>2</sup>H<sub>1</sub>]glycine (optical purity, 96%) was converted (0.5—1.0 g scale) into (R)-[<sup>2</sup>H]bromoacetic acid  $(m/e \ 139.141; M^+)$  (retention) and the latter reduced to  $(2S)-[{}^{1}H_{1}, {}^{2}H_{1}, {}^{3}H_{1}]$ ethanol with lithium aluminium tritide (inversion) as shown in Scheme 2. Without isolation,



SCHEME 2. i, NaNO, KBr, H+, 0 °C, 1 h, 5%; ii, LiAlT, (50 mCi mmol<sup>-1</sup>); iii, Cr<sub>2</sub>O<sub>7</sub><sup>2-</sup>.

chromic acid oxidation of the evaporated, ethereal extract from this reduction furnished a specimen of chiral acetic acid whose configuration was determined by the coupled enzyme assay<sup>1,2</sup> to be (R)-[<sup>2</sup>H<sub>1</sub>,<sup>3</sup>H<sub>1</sub>] (92% optical purity measured on a 1 mCi mmol<sup>-1</sup> sample). The use of these samples now easily available in 100-500 mg quantities in porphyrin and corrin biosynthesis is under investigation. We thank N.I.H. for support of this work.

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